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Silk gland morphology of the net-casting spider Deinopis spinosa

Sandra M Correa-Garhwal (Scorrea-garhwal@amnh.org)

American Museum of Natural History https://orcid.org/0000-0002-6889-2688

Jay Stafstrom Cornell University

Richard H. Baker

American Museum of Natural History

Cheryl Y. Hayashi

American Museum of Natural History

Ronald Hoy

Cornell University

Kern Reeve

Cornell University

Jessica E. Garb

University of Massachusetts Lowell, Lowell

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1 Silk gland morphology of the net casting spider *Deinopis spinosa*

- 3 Sandra M. Correa-Garhwal^{1*}, Jay Stafstrom², Richard H. Baker¹, Cheryl Y. Hayashi¹, Ronald
- 4 Hoy², Kern Reeve², & Jessica E. Garb³
- 5 ¹Division of Invertebrate Zoology and Institute for Comparative Genomics, American Museum
- 6 of Natural History, New York, NY 10024, USA
- 7 ²Department of Neurobiology and Behavior, Cornell University, Ithaca, NY 14853, USA
- 8 ³Department of Biological Sciences, University of Massachusetts Lowell, Lowell, MA 01854,
- 9 USA
- 10
- 11 *Corresponding author
- 12 Email addresses:
- 13 *SMCG: <u>scorrea-garhwal@amnh.org</u>
- 14 JS: js2627@cornell.edu
- 15 RHB: rbaker@amnh.org
- 16 CYH: chayashi@amnh.org
- 17 RH: <u>rrh3@cornell.edu</u>
- 18 JRG: jessica_garb@uml.edu
- 19 KR: hkr1@cornell.edu
- 20

21 Abstract. Net-casting spiders (Deinopidae) are cribellate spiders that spin a rectangular, sticky 22 net that is held stretched between the claws of their first two pairs of legs. Deinopids produce 23 eight distinct silk types, but knowledge of the silk-producing morphologies is mostly limited to 24 the spigots associated with different fibers. As there have been no studies of deinopid silk gland 25 structure, we dissected all the silk glands from Deinopis spinosa and document their number and morphology. We found silk gland position and morphology consistent with the type and number 26 27 of silk spigots described for Deinopidae. Notably, for the first time, we describe the silk glands associated with cribellate silk: paracribellate, pseudoflagelliform, and cribellar silk glands. Our 28 findings support the homology of pseudoflagelliform glands with araneoid flagelliform glands 29 30 and will have importance for informing our understanding of spider web evolution. 31

32 Keywords: Deinopid, cribellate silk, silk glands, spigot, spinneret

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Net-casting spiders (Deinopidae C. L. Koch, 1851) are skilled nocturnal hunters. These cribellate 34 35 spiders spin an unusual type of orb-web: a rectangular, sticky net that the spider holds stretched 36 between the claws of its first two pairs of legs. Suspended, the spider hovers motionless at night, then when prey is detected, it rapidly expands its net and flings it over prey moving on the 37 38 substrate beneath them or flying above them (Coddington & Sobrevila, 1987). Studies on 39 Deinopis spinosa Marx, 1889 (Marx, 1889) have shown that this unique foraging strategy 40 involves multiple specializations, including extremely sensitive night-vision to capture prey off 41 the ground (Stafstrom & Hebets, 2016) and extremely extensible silk (Blackledge & Hayashi, 42 2006).

43

44 Deinopids use a dry, Velcro-like adhesive in their prey-capture web, composed of cribellar silk 45 surrounding a pair of supporting fibers, a pair of undulating fibers, and a mat of connecting 46 fibers. Cribellar silk is produced from a specialized plate called the cribellum. The cribellum is 47 densely covered with thousands of miniature spigots from which nanofibrils emerge and then are 48 teased into highly-coiled, cloudy masses of fibers. The supporting fibers are produced in the 49 pseudoflagelliform glands, the undulating fibers in the minor ampullate silk glands and the connecting fibers in the paracribellate silk glands (Peters, 1984, 1992). While the external 50 51 spinning apparatus (spinnerets and their spigots) has been characterized in a few works

52 (Coddington, 1989; Griswold et al., 1998, 2005; Murphy & Roberts, 2015; Peters, 1992), to our 53 knowledge, there are no previous morphological studies of the silk glands associated with 54 deinopid spigots.

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56 Here, we document all silk gland types present in the net-casting spider D. spinosa and describe 57 their position, size, and morphology, drawing comparisons with current knowledge about their 58 spigots, as well as silk gland morphology outside of deinopids. Our findings are consistent with the number and variety of spigot morphologies and for the first time for any spider, we describe 59 60 the morphology of the silk glands used in the cribellate capture silk.

61

Mature female D. spinosa were collected in Florida (2021 and 2022). All spiders were 62 anesthetized with CO₂ and dissected under a stereomicroscope with forceps in saline sodium 63 64 citrate buffer. Images of all tissues were collected from two individual females using a ZEISS Axiocam 105 Color Microscopy Camera mounted on a ZEISS 435063-9010-100 Stemi 305 65 66 Stereo Microscope with the ZEN Blue software. We identified eight types of silk glands: 67 aciniform, cribellar, major ampullate, minor ampullate, paracribellate, pseudoflagelliform, pyriform, and tubuliform (Fig. 1), which correspond to the eight types of silk spigots described 68 69 for deinopid spiders (Fig. 2) (Coddington, 1989; Griswold et al., 1998; Murphy & Roberts, 2015; 70 Peters, 1992). Each gland type was confirmed by tracing its duct to the spinneret on which it 71 terminated. An additional mature female was anesthetized with CO₂ and the abdomen was 72 severed at $\sim 30\%$ anterior of the spinnerets. The spinnerets were prepared following (Townley & 73 Harms, 2017). In short, spinnerets were submerged in 2X SDS-PAGE running buffer 74 (ThermoFisher) for 3 days, then dehydrated in an ethanol series: 30%, 50%, 70%, 85%, 95%, 75 and then twice in 100% (24 hour each). Spinnerets were critical-point dried using a Tousimis 76 Samdri CPD, mounted on SEM stubs, sputter-coated with a Denton Vacuum Desk IV, and 77 examined in a JEOL 6390 scanning electron microscope. 78

79 The major ampullate and minor ampullate glands were astonishing in number and shape. There

80 were ten pairs of major ampullate glands (Fig. 1A), corresponding to the ten major ampullate

spigots on each anterior lateral spinneret (ALS; Fig. 2A and 2B) (Coddington, 1989; Peters, 81

82 1992). By contrast, most mature araneoids have only one pair of major ampullate glands

83 (Coddington 1989). Many non-araneoid spiders, such as eresids, nicodamids, desids, 84 amaurobiids, agelenids, and lycosoids, have been described as having more than one pair of 85 major ampullate silk spigots as mature individuals (Griswold et al., 1998, 2005; Řezáč et al., 2017). Similarly, most juvenile spiders have multiple major ampullate spigots which have been 86 87 proposed to be used alternatively during molting. That is, one spigot functions in even-numbered instars and the other in odd-numbered instars (Townley et al., 1993). The silk glands associated 88 89 with these spigots also undergo changes during ecdysis: larger ampullate glands (usually those that stay through maturity) become non-functional pre-ecdysis while the smaller ampullate silk 90 glands become functional, thus allowing for continuous use of ampullate silk. 91

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93 We also found the elongated D. spinosa major ampullate glands to vary in size within an 94 individual, a phenomenon rare among araneid and theridiid spiders (Berger et al., 2021; Chaw & Hayashi, 2018; Coddington, 1989). However, such variation has been described for some 95 agelenid and cribellate amaurobiid species (Řezáč et al., 2017). In the case of agelenids, (Řezáč 96 97 et al., 2017) showed that they have a system of three pairs of glands, in which by following the 98 Townley et al. (1993) model, each silk gland will function at a different time during the molting 99 cycle. In general, major ampullate silk glands have three distinct parts: a secretory tail (Fig. 1A, 100 star), an ampule-shaped storage sac, and an elongated spinning duct (Fig. 1A, arrow) (Chaw & Hayashi, 2018; Vollrath & Knight, 1999). In D. spinosa, all major ampullate silk glands have an 101 102 ampule-shaped sac and an elongated spinning duct, with the three longest glands (Fig. 1A, left group) also having a long tail similar to other species (Chaw & Hayashi, 2018; Clarke et al., 103 104 2017; Vollrath & Knight, 1999). The other seven major ampullate silk glands have shorter tails, 105 tend to be clustered together, and are smaller, approximately one-fourth the size of the longest 106 glands (Fig. 1A, right group). We also found differences in the size of the ampullate spigots on 107 the ALS (Fig. 2A) associated with these glands: there are three larger spigots on the inner side 108 closer to the pyriform spigots and a cluster of seven smaller major ampullate spigots on the outer 109 side (Fig. 2B). Given the presence of additional glands in juvenile spiders (data not shown), the 110 current understanding of molt-related changes in silk glands (Townley et al., 1993), and our 111 observation that all major ampullate glands are filled with silk dope, we posit that all ten major ampullate silk glands found in mature D. spinosa are fully functional. Whether the 112

morphological differentiation in *D. spinosa* major ampullate silk glands translates into usage or
functional diversification of major ampullate fibers is unclear.

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116 Historically, minor ampullate silk glands were named due to their morphological similarity to 117 major ampullate silk glands, albeit smaller in size. In D. spinosa, the minor ampullate silk glands 118 were not found to be ampule-shaped, as in other orb-weaving spiders, but instead were identified 119 based on their location, by following their ducts to the minor ampullate spigots on the anterior 120 median edge of each posterior median spinneret (PMS; Fig. 2A and 2D). We observed only one 121 minor ampullate spigot on each PMS. Moreover, we found no evidence of a second pair of minor 122 ampullate spigots. By contrast, Peters (1992) found two ampullate spigots in Deinopis subrufus 123 (Koch, 1878): a large one corresponding to minor ampullate and a smaller one that was difficult to observe. Following Peters (1992), Griswold et al. (2005) reported Deinopis as having two 124 125 minor ampullate spigots, although only one minor ampullate spigot was visible in their 126 micrographs of Deinopis. It may be that D. spinosa and D. subrufus differ in the number of 127 minor ampullate silk glands. In D. spinosa, minor ampullate silk glands are bifurcated in the sac 128 (or secretory section), have a very long secretory duct, and a short tail compared to major 129 ampullate silk glands (Fig. 1B). Bifurcation of minor ampullate silk glands has also been observed exclusively in other cribellate spiders (Řezáč et al., 2017), suggesting that bifurcation 130 131 could be functionally paired with the production and/or function of the undulating fibers in cribellate silk. 132

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134 The other silk glands that do not have any function in cribellate capture threads are the 135 aciniform, pyriform, and tubuliform (also known as cylindrical) silk glands. We found the 136 aciniform and pyriform silk glands to be among the smallest gland types (Fig. 1C, D). The 137 aciniform silk glands lack morphologically distinct tails and were tightly packed in a grape 138 cluster-shaped formation (Fig. 1C). There were two pairs of clusters (one on each side): a cluster 139 of ~47 aciniform glands attached to the posterior lateral spinneret (PLS) and a cluster of ~96 140 aciniform silk glands attached the PMS (Fig. 1C, D). Moreover, morphological differences in 141 aciniform silk glands and spigots in *D. spinosa* were not observed as with other spiders (Kovoor 142 & Peters, 1988; Peters & Kovoor, 1980).

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144 Pyriform silk glands were also tightly clustered, with ~100 glands arranged in a "baseball-glove" 145 shape (Fig. 1D). It was not possible to separate individual aciniform or pyriform silk glands 146 without destroying them. Both aciniform and pyriform silk glands were found in the hundreds 147 and have very small and thin ducts. Tubuliform silk glands were long, with a yellow-orange 148 color, and take on the previously described noodle-shape of tubuliform glands from other Entelegynae species (Fig. 1E). We found the tubuliform silk glands to be extraordinarily 149 150 numerous, ~90 pairs, compared to araneoid spiders, which possess only three pairs of tubuliform 151 glands (Coddington, 1989; Griswold et al., 1998, 2005; Murphy & Roberts, 2015). Deinopids 152 construct very densely woven and hard egg cases (Barrantes et al., 2014), and these properties 153 likely result from their large number of tubuliform glands.

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Prey capture silk in deinopids is a composite of multiple components, including the ultrafine 155 156 cribellar fibrils, the extensible core pseudoflagelliform silk fibers and connective paracribellate 157 fibers. A fourth component of deinopid cribellate silk is minor ampullate silk which is used in the undulating fibers as mentioned above. The morphologies of the glands associated with these 158 159 three silks are largely unknown for any species. We identified a single noodle-shaped silk gland 160 attached to a single large spigot present on the lateral edge of the PLS (Fig. 2A and 2C). This 161 spigot has been considered to be homologous to the flagelliform silk spigots of ecribellate orb-162 web weavers because of similarity in position and shape and was thus called the 163 pseudoflagelliform spigot (Alfaro et al., 2018; Coddington, 1989; Peters, 1992). It follows that 164 the silk gland attached to this spigot is the pseudoflagelliform silk gland (Fig. 1F). Like the 165 flagelliform silk gland, the pseudoflagelliform silk gland is elongated ampule-shaped, with a distinct tail and a small kink towards the duct (Fig. 1F). As for the second component of 166 167 cribellate silk, the paracribellate silk glands were identified by tracing the glands to the 168 corresponding spigots on the PMS (Fig. 2D). The paracribellate silk glands are ampule-shaped 169 with thin elongated ducts and short tails. Paracribellate glands are numerous ~ 120 (~ 60 silk 170 glands on each side), tightly arranged in clusters (Fig. 1G), and separating them was challenging 171 (but see Fig. 1H). The final silk type, cribellar silk glands, are attached to a special plate-like 172 organ called the cribellum (Fig. 1I, brown line), which is an identifying feature of cribellate 173 spiders. The cribellar silk glands were so snugly clustered together that it was impossible to 174 isolate a single gland (Fig. 11). The number of glands was also so vast that they were impossible

175 to count. However, a study of D. subrufus estimated that there were $\sim 25,000$ spigots on the 176 cribellum, suggesting ~25,000 cribellar silk glands (Peters, 1992). Histological studies shows 177 that cribellar glands are arranged as a compact mass widespread above and around the PLS and PMS (Kovoor & Peters, 1988) which is consistent with the morphology and the number 178 179 described in this work. When the cluster of D. spinosa cribellar glands was torn apart, thin ducts 180 were observed and the glands appeared to be ovoid, with no visible tails. The glands were 181 arrayed in tightly packed rows that folded back on themselves, almost giving the appearance of a box pleated skirt. 182

183

184 We show for the first time the morphology of the complete complement of silk glands from a 185 cribellate spider. Moreover, we describe the previously unknown morphology of the paracribellate, pseudoflagelliform, and cribellar silk glands. Confirmation of the presence of a 186 187 pseudoflagelliform gland in a deinopid, exhibiting morphological and positional similarity to 188 araneoid flagelliform glands has special significance for our understanding of spider silk 189 evolution and the origin of the iconic orb-web. Early debates centered on whether the orb-web 190 evolved convergently in different spider lineages or if it had a single origin (Shear, 1986). Coddington (1989) proposed homology of the pseudoflagelliform and flagelliform silk spigots, 191 192 implying that cribellate spiders spinning orb-webs and orb-web derivatives (Uloboridae and 193 Deinopidae) were the sister group of Araneoidea (ecribellate orb-weavers), and that the orb-web 194 traced to a single origin in their most recent common ancestor. However, recent phylogenomic 195 reconstructions of the spider tree of life are contentious: several have repositioned deinopids, 196 uloborids, and araneoids in different ways; and they disagree as to whether the orb-web had a 197 single, but more ancient origin (e.g., Coddington et al., 2019; Garrison et al., 2016), versus 198 multiple, convergent origins (e.g., Fernández et al., 2018; Kallal et al., 2020). Adding to this, 199 (Eberhard, 2022) recently argued that a suite of several web-building behaviors shared across 200 cribellate and ecribellate orb-weavers are homologous, and strongly favors a single origin of this 201 web architecture. Like these behavioral characters, the positional and morphological similarity of 202 the pseudoflagelliform and flagelliform silks glands in deinopids and araneoids, reinforces that 203 they are homologous (Alfaro et al., 2018; Correa-Garhwal et al., 2022). Given that silk extruded 204 from these glands is predominantly used in the orb-web capture spiral (for both cribellate and 205 ecribellate orb-weavers) or in deinopid capture silk supporting fibers, resolution of the

206 phylogenetic positions of these spider lineages will not only inform the number of times the orb-

- 207 web evolved, but also the number of times the pseudoflagelliform/flagelliform glands have been
- 208 lost, as well as how silk fibers extruded from their spigots have functionally transformed over
- 209 evolutionary time. Beyond the importance of the pseudoflagelliform gland, future studies are
- 210 needed to understand the function and evolution of the entire suite of silk glands that allow
- 211 deinopids to capture prey with their remarkable, hyper-extensible capture nets.
- 212

Figure 1. Silk glands of the net-casting spider *Deinopis spinosa*. Silk glands are oriented with

the ducts (arrow) up and tails (star) at the bottom. A. Major ampullates, B. Minor ampullate, C.

215 Aciniform cluster, D. Pyriform cluster, E. Tubuliform single, F. Pseudoflagelliform, G.

216 Paracribellate cluster, H. Paracribellate single, and I. Cribellar cluster. Major ampullate, minor

ampullate, aciniform cluster, pyriform cluster, pseudoflagelliform, and paracribellate are paired

218 inside the spider and only one side is depicted. Scale bar = $500 \ \mu m$.

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220 Figure 2. Cribellum and spinnerets of adult female *Deinopis spinosa*. A. Overview of spinnerets

B. Anterior Lateral Spinneret (ALS), C. Posterior Lateral Spinneret (PLS), D. Posterior Median

222 Spinneret (PMS) E. Cribellum, with close-up inset on upper left. Abbreviations as follows: Cr

cribellum and spigot types, Py: pyriform, Ma: major ampullate, Tu: tubuliform, Ac: aciniform,

- 224 Pflag: pseudoflagelliform, Mi: minor ampullate, Pcr: paracribellate.
- 225

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- 230

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